REVIEW

Antenatal risk factors, cytokines and the development of atopic disease in early childhood

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Atopic diseases are complex entities influenced by an array of risk factors, including genetic predisposition, environmental allergens, antenatal exposures, infections and psychosocial factors. One proposed mechanism by which these risk factors contribute to the development of atopic disease is through changes in the production of T helper cell type 1 (Th1) and T helper cell type 2 (Th2) cytokines. The objectives of this review are to discuss antenatal exposures that are associated with paediatric atopic diseases, to discuss the influence of the intrauterine environment on neonatal immune responses, to provide an overview of the Th1 and Th2 pathways and how they relate to atopic disease, and to summarise our current understanding of the association between cytokine responses in cord blood and the development of atopic disease in early childhood.

ith the recent rise in atopic disease prevalence, it would be beneficial for clinicians to become familiar with research advances made in the area of paediatric atopic disease pathogenesis. Atopic diseases—asthma, allergic rhinitis, atopic dermatitis and food allergy—are complex entities with an array of risk factors that may be categorised into genetic predisposition, environmental allergens, antenatal exposures, infections and psychosocial factors (fig 1).

Genetic predisposition is central to the development of atopic disease as shown by the increased disease prevalence among first-degree relatives of affected people and those with a positive family history of atopic disease, ¹⁻³ by monozygotic versus dizygotic twin studies, ⁴ and by the identification of numerous chromosomal linkages, single-nucleotide polymorphisms and haplotypes that are associated with an increased risk of atopic disease or biomarkers of atopy, such as serum IgE levels. ⁵⁻⁷ Atopic diseases are inherited as complex diseases involving the interplay of as many as 20 separate genes. Evidence of gene—environment interactions shows that the environment has a modifying effect on the expression of certain genes in atopic disease ^{8 9}

The environment and infectious diseases affect the development of atopic disease, and have recently received a great deal of attention in view of the recent upsurge in atopic disease prevalence. It is unclear whether infections alter actual disease risk; however, respiratory syncytial virus and rhinovirus infections are associated with an increased likelihood of subsequent wheezing and childhood asthma.10 11 Supporters of the controversial "hygiene hypothesis" attribute the increased prevalence of atopic disease in the Western world to a relative decrease in infectious diseases associated with trends that include, but are not limited to, smaller family size, an increased emphasis on hygiene and the widespread use of antibiotics.12 13 This theory is supported by evidence showing a decreased risk of atopic disease where there is an increased exposure of young children to microorganisms, including an increased exposure to endotoxin in the first several months of life,14 daycare attendance in infancy,15 living with older siblings, 12 15 living on a farm 16 and early pet exposure.17 18 Additional studies, but not all,19 have shown an association between antibiotic use in early life and an increased risk of asthma or atopy later in childhood.20 21

Paediatricians are generally familiar with genetic predisposition and many of the postnatal exposures associated with atopic disease. However, antenatal exposures associated with the development of atopic disease and recent advances in atopic disease pathogenesis may not be in the purview of the general paediatrician or practitioner. This review focuses on the role of the intrauterine environment and antenatal exposures in the development of atopic disease in early childhood. The objectives of this review are to discuss antenatal exposures that are associated with paediatric atopic diseases, to discuss the influence of the intrauterine environment on neonatal immune responses, to provide an overview of the T helper cell type 1 (Th1) and T helper cell type 2 (Th2) pathways and how they relate to atopic disease, and to summarise our current understanding of the association between cytokine responses in cord blood and the development of atopic disease in early childhood.

ANTENATAL EXPOSURES ASSOCIATED WITH PAEDIATRIC ATOPIC DISEASE AND CORD BLOOD BIOLOGICAL ASSAYS SHOWING EVIDENCE OF NEONATAL ANTIGEN-SPECIFIC IMMUNITY

The importance of the intrauterine environment in atopic disease pathogenesis is supported by data showing a greater influence of maternal over paternal atopy on disease risk in the offspring^{1 2}

Abbreviations: CBMC, cord blood mononuclear cell; IFN, interferon; IL, interleukin; PBMC, peripheral blood mononuclear cell; Th1, T helper cell type 1; Th2, T helper cell type 2; TNF, tumour necrosis factor

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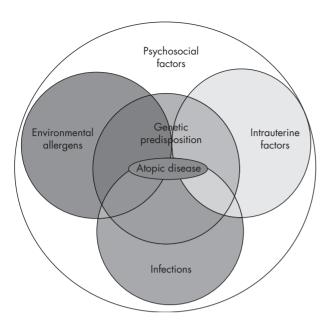


Figure 1 Atopic disease risk factors.

and multiple antenatal risk factors for paediatric atopic disease.² ^{22–24} Several maternal health characteristics and behaviours during pregnancy are associated with paediatric atopic disease in the offspring. These include low maternal parity,²⁵ respiratory and genitourinary infections,²³ ²⁶ cigarette smoking²² ^{27–29} and antibiotic use during pregnancy,³⁰ a proxy for maternal infection. At birth, risk factors for atopic disease that may reflect intrauterine exposures include higher gestational age,² low birth weight and prematurity,²² and delivery by caesarean section.^{31–33}

Beyond the epidemiological associations found between certain antenatal exposures and the subsequent development of atopic disease in offspring, there are biological measures that show evidence of neonatal T cell and B cell immune responses to antenatal exposures. Cord blood biological assays among offspring show evidence of neonatal immune responses to antigens. 34 35 At birth, in vitro allergen-stimulated T cell or cord blood mononuclear cell (CBMC) proliferation occurs in response to a variety of environmental allergens, including βlactoglobulin, birch pollen, bovine serum albumin, cat fur, cockroach, house dust mite, mouse, ovalbumin and timothy grass pollen.34 36-39 According to some investigators, these responses indicate that neonatal immune responses, thought to be naive, may in fact be influenced by antenatal exposures,³⁷ but others have challenged the specificity of such studies.⁴⁰ Antigen-specific B cell responses to maternal antigen exposures during pregnancy have also been described. After giving tetanus vaccine to pregnant women, investigators have detected antigen-specific IgM production in cord blood.41 In addition, measurable total and allergen-specific IgE in cord blood provides evidence that isotype-class switching occurs in response to in utero allergen exposure.35

Several plausible mechanisms by which in utero sensitisation may occur are seen. One possibility is that the antigen or processed peptide to which the mother is exposed during pregnancy reaches the placenta where antigen presentation occurs. In support of this mechanism, the dust mite antigen, *Dermatophagoides pteronyssinus* 1, has been detected in amniotic fluid and umbilical cord blood, introducing the possibility of transplacental passage of antigens.⁴² In addition, antigen presenting cells have been detected in the placenta and have

Cytokine	Description				
Th1 cytokines					
ΙΕΝγ	Most reliably produced by Th1 cells Th2 antagonist through activation of T cell transcription factor, T-bet and subsequent inhibition of Th2 cell differentiation Activates macrophages Stimulates production of IgG Inhibits IL4-induced IgE synthesis in human PBMCs				
TNFα TNFβ	Stimulates phagocytosis, and production of IL1 and oxidants Involved in cell proliferation, differentiation and apoptosis Cytotoxic to a range of tumour cells				
IL12	Produced by various APCs Induces IFNγ				
Th2 cytokines					
IL4	Reliably Th2 Polarises naive T cells towards the Th2 phenotype, through activation of the transcription factor GATA3 With IL13 causes isotype switching of B cells to synthesise IgE after exposure to allergen Activates and stimulates bone marrow production of eosinophils				
IL5	Activates and stimulates bone marrow production of eosinophils				
IL9 IL13	Involved in eosinophil and mast cell development With IL4 causes isotype switching of B cells to synthesise IgE after exposure to allergen in humans Contributes to airway hyper-reactivity in animals				

been shown to facilitate antigen-induced T cell proliferative responses.⁴³

peripheral blood mononuclear cell; TNF, tumour necrosis factor.

OVERVIEW OF TH1 AND TH2 CYTOKINES AND HOW THEY RELATE TO ATOPIC DISEASE

In this section, we provide a simplified overview of Th1 and Th2 cytokines and how they relate to atopic disease. In an effort to better understand disease pathogenesis and to identify biological measures to predict atopic disease and potential therapeutic modalities to treat disease, much attention has recently been focused on Th1 and Th2 cytokine production and on their counter-regulatory actions. A general understanding of Th1 and Th2 cytokines is necessary to understand how cord blood cytokines relate to paediatric atopic disease development.

Th1 and Th2 lymphocytes, thought to be the differentiated progeny of a population of naive lymphocytes, are defined by the cytokines that they produce. Table 1 describes the major Th1 and Th2 cytokines. To a large extent, Th1 and Th2 pathways—influenced by cytokines, other immunological cells and transcription factors—are thought to be counter-regulatory (fig 2). The Th1 pathway is essential for cell-mediated immunity and occurs in response to some bacterial infections. The Th2 pathway, essential for all humoral immunity, is thought to play a major part in atopic disease (fig 3), with multiple studies showing an association between the atopic phenotype and increases in Th2 cytokines in the sera and the bronchoalveolar lavages of affected people.⁴⁴⁻⁴⁶

The Th2 dominance seen in people affected by atopic disease has led many researchers to believe that environmental and infectious exposures, including antenatal exposures, may modulate disease risk through changes in the balance between the Th1 and Th2 cytokine pathways. Several exposures associated with an enhanced Th2 response and a reduced Th1 response are also associated with atopic disease. For example, cigarette smoking during pregnancy, a known risk factor for

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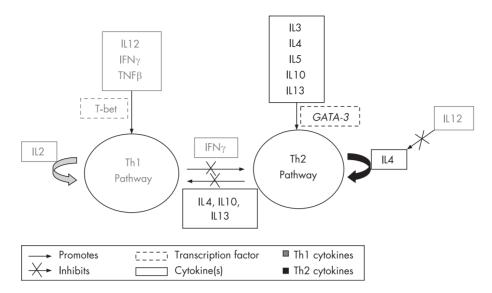


Figure 2 Overview of the counter-regulatory actions of the T helper type 1 and 2 (Th1 and Th2) pathways. IFN, interferon; IL, interleukin; TNF, tumour necrosis factor.

childhood asthma, is associated with increased levels of the Th2 cytokine, interleukin (IL)13, and decreased levels of the Th1 cytokine, interferon (IFN) γ , in cord blood.⁴⁷ Respiratory syncytial virus infection, a risk factor for wheezing as noted above, is also associated with a Th2 response.⁴⁸ Similarly, in some studies, early antibiotic use in infancy is associated with an increased risk of atopic disease²⁰ ²¹ and is also associated with a Th2-dominant response in mice.⁴⁹ On the other hand, exposures associated with an enhanced Th1 response are associated with a reduced risk of atopic disease. Specifically, Th1-inducing infections, such as *Mycobacterium tuberculosis* and hepatitis A virus, are associated with a reduced risk of atopic disease among affected people.⁵⁰ ⁵¹

The relationship between Th-1/Th-2-inducing exposures and atopic disease is not always straightforward, and some studies question these associations. Although one study showed that Th1-inducing immunisation with bacillus Calmette–Guérin was associated with a reduced risk of atopic disease, another study showed that it was not.⁵⁰ ⁵² Similarly, although Shaheen *et al*⁵³

showed that people with a history of Th-1 inducing measles infection had a lower likelihood of atopy than those without measles, a later study showed that measles infection was associated with a higher risk of atopic disease.⁵⁴ Other studies show that Th1 and Th2 pathways are not always counterregulatory. For instance, in mice, Hansen *et al*⁵⁵ showed that the production of the Th1 cytokine, IFNγ, was insufficient to counteract the effects of IL4 and IL5. Instead of attenuating Th2 cell-induced airway hyper-reactivity and inflammation, Th1 cells actually caused severe airway inflammation.

The intriguing associations between some Th-1-inducing exposures and a reduced risk of atopic disease, Th-2-inducing exposures and an increased risk of atopic disease, and the counter-regulatory actions of the Th1 and Th2 pathways for many years have provided an immunological basis for the hygiene hypothesis. In addition, the Th1/Th2 paradigm has prompted a great deal of research that has broadened our understanding of atopic disease pathogenesis. Although the relationship between cytokine responses and atopic disease is

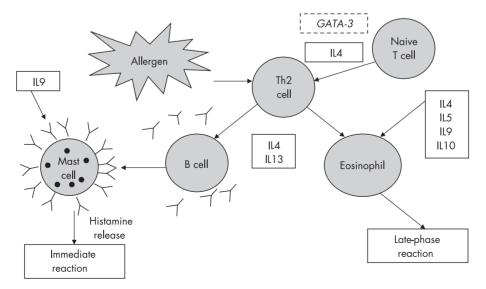


Figure 3 T helper cell type 2 (Th2) cytokines and atopic disease.

complex and not fully understood, we know that cytokines play an important part in atopic disease.

CURRENT UNDERSTANDING OF THE ASSOCIATION BETWEEN CYTOKINE RESPONSES IN CORD BLOOD AND SUBSEQUENT DEVELOPMENT OF ATOPIC DISEASE

In the search for predictive biological markers for atopic disease, investigators have recently explored the relationship between CBMC and peripheral blood mononuclear cell (PBMC) cytokine responses to mitogen and antigen stimulation and the subsequent development of atopic disease. Table 2 provides a sample of studies from major research groups around the world that have substantially contributed to this body of work. The most consistent finding from these and related studies has been that people who develop atopic disease and those who have a positive family history of atopy even in the absence of disease have lower levels of the Th1 cytokine, IFNγ, at birth when compared with their unaffected counterparts.^{3 36 39 56 57} IFNy levels produced in response to allergen-stimulated CBMCs have been shown to be inversely related to cord blood IgE levels, 56 a marker with high specificity but low sensitivity for atopic disease.

Low levels of IFN γ may represent impaired Th1 pathway function, immature development of the Th1 pathway, early destruction of Th1 cells 58 or dominance of Th2 immune

responses. Some investigators have speculated that the relative lack of Th1 cytokine expression in newborns at risk of atopic disease may be due to antigen-presenting cell immaturity and an inability to release IL12, the major induction cytokine for the Th1 pathway. The initially low levels of IFN γ seem to extend beyond the neonatal period, as children who develop atopy also lack the normal Th1 response to bacillus Calmette–Guérin vaccination in infancy. ⁵⁰ Also, children at increased risk of atopic sensitisation have an attenuated production of the Th1 cytokine, IFN γ , in early infancy. ⁵⁹

Levels of Th2 cytokines produced in response to antigen stimulation of CBMCs among children who develop atopic disease, however, are less consistent than the low levels of IFN γ that have been found. Among people at high risk of, or who develop, atopic disease, investigators have shown a Th2 dominant response as shown by increased levels of IL5,60 61 whereas others have shown lower levels of Th2 cytokines such as IL13.62

Some reports suggest that cytokine responses change during early childhood, and that such changes differ for people without atopic disease when compared with people with atopic disease. For people without atopic disease, studies have shown a decline in the Th2 cytokine IL4 and an increase in production of the Th1 cytokine IFN γ in the first 2 years in response to house dust mite antigen-stimulated mononuclear cells.⁶³ At the same time, children who develop atopic disease show an

Description of study		Methods and measures				Major findings related to cytokines and atop
Source	Study population	Age: blood sample	Cytokines	Allergens/mitogens	Major clinical outcomes	. disease
Contreras et al ⁵⁷	112 children all with a parental history of asthma or allergy (54 w/and 58 w/o atopic disease) Birth to age 2 years	Age 2 years: PBMCs	IFNγ TNFα IL10 IL13	Bla g 1 (cockroach) Der f 1 (HDM) Feld 1 (cat)	Atopic disease: Doctor/nurse diagnosed Eczema Hayfever Allergic rhinitis Inhalant allergy Repeated wheeze	Children w/ atopic disease had lower IFN _Y (Th1 cytokine) levels in response to HDM an cockroach allergen stimulation of PBMCs the non-atopic children. This finding appeared to be more pronounced for children w/atopic disease and repeated wheeze.
Prescott et al ^s	60 children (44 w/positive family history of atopy; 16 w/o family history) Birth to age 6 years, every 6 months All born by elective caesarean section	Birth: CBMCs Age 6 months: PBMCs Age 12 months: PBMCs Age 18 months: PBMCs Age 24 months: PBMCs	IFNy IL4 IL5 IL6 IL9 IL10 IL13	HDM Ovalbumin Fel d 1 (cat) PHA Tetanus toxoid	Allergic (atopic) disease: Doctor-diagnosed asthma Asthma: recurrent wheezing (≥3 episodes) Doctor-diagnosed eczema SPT at age 6 years	Children w/family history of allergy had low IFN ₇ (Th1 cytokine) responses to PHA stimulation of CBMCs (at birth) compared w children w/o a family history. Children with atopic disease at 6 years had increase in the 1st 2 years of life in IL5 (Th2 cytokine) mRNA in response to HDM, but the was no change in the non-atopic group. Positive SPT to HDM at 6 years was associat w/ higher IL13 (Th2 cytokine) responses to HDM at 1 year.
Neaville et al ⁶³	285 children Birth to age 1 year Neonates w/at least one parent w/ respiratory allergies or asthma	Birth: CMBCs Age 1 year: PBMCs	IFN ₇ IL5 IL10 IL13	РНА	Atopic dermatitis Food allergy Antigen-specific IgE levels	Lower IL10 (Th2 cytokine) production in response to PHA-stimulated CBMCs (at birth was a risk factor for egg sensitisation at 1 ye of life. For the cohort, IL5 (Th2 cytokine) response increased while IFN γ (Th1 cytokine) decreas over the 1 year of life
Kondo et al ^{ss}	21 children (7 w/ allergic disorder and 14 w/o) Birth to age 6 years Full term, vaginally born infants	Birth: CBMCs	IFN _γ IL2	Ovalbumin Bovine serum albumin	Allergic disorder: Atopic dermatitis Bronchial asthma Allergic rhinitis Food allergy	Children who developed allergic disorder by age 6 years had lower maximal concentration of IFNγ (Th1 cytokine) in response to ovalbumin or bovine serum albumin stimulation of CBMCs than those w/o allergic disorder Maximal concentrations of IL2 did not differ between children w/ and w/o allergic disorder

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upregulation of Th2 cytokines, including IL5,62 IL9 and IL13, in response to allergen or mitogen stimulation of PBMCs in the first 2 years of life, 64 but the exact timing of this Th2 skewing is unknown.

Some evidence shows that most of the tested newborns, independent of their risk of atopy, have Th2-skewed responses to common environmental allergens, with cord blood elevations of cytokines, including IL4, IL5 and IL13.64 This Th2 skewing is thought to reflect the Th1/Th2 state of the mother, as women towards the end of pregnancy are believed to be Th2 dominant as a means of maintaining the pregnancy and protecting the fetus against the toxic effects of Th1 cytokines, such as IFNγ.65 Lending strength to this idea, some recent studies have shown a correlation between maternal and neonatal cytokine profiles.66 67 Some think that people without atopic disease shift from being Th2 skewed at birth to being more Th1/Th2 balanced, and that people with atopic disease instead show an upregulation of Th2 and continue to be Th2 skewed thereafter. Whether the ultimate cytokine profiles of an individual are influenced by antenatal or postnatal exposures or a combination of the two is unknown.

This review would be incomplete without mentioning some of the limitations to the current literature on cord blood cytokines and their relationship to paediatric atopic disease. Research in this area is limited by the operational definition of disease, with some investigators comparing cytokine profiles of children with and without atopy. Children with atopy are defined by a positive family history, positive skin-prick testing or radioallergosorbent testing, or some combination of these, rather than actual symptomatic, clinical disease. Inherent limitations regarding the techniques used for measuring cytokines also exist. For example, certain cytokine levels, including IL4 and IL9, and at times IFNy, are often below detectable limits in cord blood by standard ELISA methods and have, therefore, been based on semiquantitative reversetranscriptase polymerase chain reaction testing for specific mRNA. CBMCs and PBMCs may not show the immunological cells found in the airways or lungs, and cytokine responses from these cells may not reflect those in the target tissues.

CONCLUSIONS

Rapid progress has been made in our understanding of how neonatal cytokine production relates to the subsequent development of atopic disease; however, the exact determinants of neonatal cytokine profiles are not well understood. Studies that relate antenatal sociodemographic and health conditions to cord blood cytokine profiles are just beginning. Increasingly, we have begun to not only recognise the importance of antenatal exposures in the subsequent risk of atopic diseases but also postulate the mechanisms by which such relationships occur. In particular, cytokine profiles in cord blood appear to alter in response to antenatal exposures and may predict later development of allergic disease. Although children with atopic disease seem to show a Th2 dominance once they are diagnosed with allergic disease, it is not clear when this Th2 dominance develops; however, it is believed to develop in early childhood. Despite limitations that affect the overall generalisability of work in this area, an increasing prevalence of atopic disease, along with growing evidence that antenatal factors contribute to disease development, calls for further research in the area of neonatal cytokines and their effect on the development of childhood atopic disease.

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